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HLA Desensitization in Solid Organ Transplantation: Anti-CD38 to Across the Immunological Barriers

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Transplantation

Abstract

 The presence of anti-human leucocyte antigen (HLA) antibodies in the potential solid organ transplant recipient's blood is one of the main barriers to access to a transplantation. The HLA sensitization is associated with longer waitlist time, antibody mediated rejection and transplant lost leading to increased recipient's morbidity and mortality. However, solid organ transplantation across the HLA immunological barriers have been reported in recipients who were highly sensitized to HLA using desensitization protocols. These desensitization regimens are focused on the reduction of circulating HLA antibodies. Despite those strategies improve rates of transplantation, it remains several limitations including persistent high rejection rate and worse long-term outcomes when compare with non-sensitized recipient population. Currently, interest is growing in the development of new desensitization approaches which, beyond targeting antibodies, would be based on the modulation of alloimmune pathways. Plasma cells appears as an interesting target given their critical role in antibody production. In the last decade, CD38-targeting immunotherapies, such as daratumumab, have been recognized as a key component in the treatment of myeloma by inducing an important plasma cell depletion. This review focuses on an emerging concept based on targeting CD38 to desensitize in the field of transplantation.

Introduction

HLA sensitization and antibody-mediated rejection

 Solid organ transplantation (SOT) has become the best therapeutic option for end-stage organ disease but faces two major issues: the limited transplant supply and the poor long-term transplants outcome which have not improved over the past 30 years (1–3). This observation is related to the occurrence of antibody-mediated rejection (ABMR) which remains the death-censored leading cause of transplant loss across all solid organ transplants (3,4). ABMR is defined on the association of histologic lesions (microvascular inflammation), histologic evidence of alloantibodies – endothelium interaction (c4d staining) and circulating donor-specific antibodies mostly directed against human leucocyte antigens (HLA) (3–10). Following blood transfusion, pregnancy or previous graft failure, candidates for organ transplantation can become sensitized against HLA and produce circulating anti- HLA antibodies (11,12). In particular, pending on their properties donor-specific anti-HLA antibodies (DSA), are responsible for ABMR leading to allograft dysfunction and graft loss (13– 18). Currently, immunomotoring of the transplant candidate's is routinely performed in order to stratify the immunological risk by determining the presence and specificity of anti-HLA antibodies and potential DSA (11,16). The highly sensitized patients have longer waitlist times with significant adverse effect on both quality and quantity of life (1,2). Several strategies are applied to limit the time on the waiting list of highly immunized patients such as prioritization in transplant's access, promotion of transplantation from living-donor allografts, development of kidney paired donation and desensitization.

Desensitization and solid organ transplantation's outcome

 Current desensitization strategies have been developed in kidney transplantation and extended to other solid organ transplantation (17–21). The goal of desensitization regimens in presensitized transplant candidates is twofold including the reduction of anti-HLA level to allow transplantation and the improvement of transplantation outcome through the prevention of ABMR (22). A stepwise approach is commonly used to desensitize including, (i) either high-dose intravenous immunoglobulin (IVIG) or low dose IVIG in association with plasmapheresis to remove antibodies and, (ii) anti-CD20 targeting agent, such as rituximab, to prevent rebound antibodies development by B cell depletion (23–27). Regarding the kidney transplantation field, despite the desensitizing effect, the subsequent transplantation is associated with higher rate of rejection and higher rate of hospital readmission after transplantation (28–30). However, long term outcomes for patient and graft survival have been reported to be similar to that of non-sensitized patients (31). Furthermore, the benefit of desensitization compared to remaining on the transplant waiting list has been evaluated only in few large studies and their results remain controversial (32,33). Montgomery *et al.* and Orandi *et al.* reported a survival benefit at five years after kidney transplantation in 211 and 1025 desensitized patients respectively compared to patients remaining on the waiting list (34,35). Interestingly, in a study performed on 213 desensitized recipients of living donor transplants, Manook *et al.* showed that desensitization was not associated with a survival benefit compared to matched sensitized control patients who were waitlisted (36). On the other hand, keeping patients a long time on dialysis represent a considerable financial burden while decreasing the quality and length of life for affected patients (32,33).Thus, it appear as necessary to develop novel therapeutic approaches in order to prevent ABMR and improve long-term survival of transplanted organs in highly immunized recipient.

Desensitization regimens targeting plasma cells

 The available therapeutic tools to manage the humoral response appears modestly successful in the context of SOT and alloimmunity. Indeed, antibody rebound due to plasma cells (PC), which

 do not express CD20, limit the efficacy of the most commonly used strategy combining IGIV, plasmapheresis and B cell depletion by anti-CD20 depleting agent. Targeting PC with new pharmacological tool from autoimmunity and cancer research could allow a better management of the humoral response in desensitization protocols (37). In the germinal center, after the enhancement of alloantigen responses by T follicular helper (Tfh), activated B cells develop into memory-B cells, progress to plasmablasts and ultimately to andibody-producing PC (38,39). These PC are the long- lived mediators of lasting humoral immunity and persist in medullary niche where they can secrete high-affinity complement-activating DSAs (38,40). Several emerging strategies aim to deplete PCs compartment in order to prevent ABMR (37,41). First, Interleukin 6 (IL-6) is a cytokine promoting Tfh and enhancing the progression of B cells to high-affinity antibodies producing PC (42). Tocilizumab, a first-in-class humanized monoclonal antibody (mAb) with specificity for IL-6R, reduce inflammation within the allograft during ABMR in heart and kidney transplantation (43) and induce circulating DSA reduction (44). Clazakizumab is a humanized IgG1 mAb with specificity for IL6 which can also induce circulating DSA reduction (45). Both Tocilizumab and Clazakizumab are pharmacological agents with major interest in the development of desensitization strategies targeting PC (37,46). On another hand, proteasome inhibitors represent one of the most promising solution to deplete PC in the setting of desensitization, targeting more selectively PCs population. Bortezomib and carfilzomib have been evaluated in desensitization trials, lacking control group, leading to controversial results (47,48). Both induce significant PCs depletion whereas DSA level did not significantly decrease or rebound occurred rapidly. In fact, targeting PC may lead to rapid germinal center activation by deleting the negative feedback usually provided by PC and rebound humoral immunity and compensation (49). Therefore, dual targeting approach (combining PCs depletion with proteasome inhibitors and costimulation blockade) may silence the germinal center and prevent humoral compensation. This strategy has been recently evaluated using carfilzomib and belatacept as desensitization in highly sensitized non-human primate model with a reduction of bone marrow PC, DSA levels reduction, and prolongation of allograft survival. Most animals experienced ABMR with humoral-response rebound, suggesting desensitization must be maintained after transplantation using ongoing suppression of the B cell response (50,51). An emerging therapy to induce DSA reduction and to prevent rebound DSA development is the use of antiplasma cell therapies such as anti-CD38, anti-CD19 or bispecific anti-CD3 / anti-BCMA (B cell maturation antigen). In this review, we propose to focus on anti-CD38 as a desensitization regimen in SOT.

CD38-Targeting strategies

CD38 and CD38-targeting antibodies

 The protein CD38 is a type II transmembrane glycoprotein known as a multifunctional molecule. CD38 play dual roles as receptors and ectoenzymes (52). The CD38/CD31 interactions are crucial to leukocyte adhesion and transmigration through the endothelium (53). CD38 is also an enzyme that catalyzes several reactions leading to the regulation of cytoplasmic calcium fluxes and a wide range of others physiological functions such as cellular metabolism (52). CD38, found throughout the immune system especially natural killer and PC, is highly expressed in multiple myeloma cells (54). Altogether, this has triggered the development of several CD38 antibodies to treat multiple myeloma (54–56). Daratumumab (DARZALEX®, Janssen), fully human IgG1-kappa, was the first CD38 antibody that was recognized as an emerging therapy against myeloma in the last 122 decade (57). Daratumumab have multiple effects including Fc-dependent immune-effector
123 mechanisms and direct effects. The Fc-dependent immune-effector mechanisms include antibodymechanisms and direct effects. The Fc-dependent immune-effector mechanisms include antibody- dependent cellular cytotoxicity, antibody-dependent cellular phagocytosis, and complement- dependent cytotoxicity (54,55). Direct effects include induction of apoptosis, as well as inhibition of CD38 ectoenzyme function, which may lead to disruption of the PCs niche. Those Fc-dependent 127 effects and direct effects are associated with deep and sustained $CD38⁺$ cells depletion, mostly PC and NK cells (54,55,58). The ability of daratumumab to efficacy deplete PCs compartment permit to use it as an new agent in therapeutic armamentarium for multiple myeloma (56). Large clinical trials have demonstrated significant improvements in the outcome of patients with relapsed multiple myeloma with use of daratumumab and it has been recently approved in front-line regimens (56–60). Isatuximab (SARCLISA®, Sanofi) is a chimeric IgG1-kappa which has stronger direct effects than daratumumab but lower ability to induce Fc-dependent immune-effector mechanisms, while it remains unknown whether these functional differences observed between different CD38 antibodies affect their therapeutic utility (55,61). Many other strategies targeting CD38 are under development and a selection is listed in Table 1. The CD38-targeting antibodies generally represent a safe treatment. Indeed, the most reported toxicity is infusion related reactions which remain successfully controlled by premedication and infusion rate management with low frequency of recurrence during subsequent injections (62). A higher rate of viral infections in patients treated with daratumumab has been reported in some studies leading to a recommended administration of valaciclovir during the administration of anti-CD38 antibodies (62).

Immunomodulatory effects of CD38-targeting antibodies

 CD38-targeting antibodies have immunomodulatory effects such as improving the host-anti- tumor immune response (63). Krejcik *et al.* showed that daratumumab monotherapy against myeloma was associated with both CD4+ and CD8+ T cell expansion(64). This increase in T-helper cells and cytotoxic T-cell was associated with functional modification including elevated antiviral and alloreactive functional responses, and significantly greater increases in T-cell clonality as measured by T-cell receptor sequencing (63,64). These modifications are associated with depletion of $CD38⁺$ immunosuppressive cells including regulatory T cells, regulatory B cells, and myeloid-derived suppressor cells. It is well known that such regulatory cells inhibit the host-anti-tumor immune response in the context of several malignancies including multiple myeloma (65–67). Altogether, this immunomodulatory activity of CD38 antibodies may be essential to their therapeutic efficacy. Indeed, it has been highlighted in clinical trials showing that expansion of effector T-cells and eradication of immune suppressors cells by daratumumab used against refractory and newly diagnosed multiple myeloma was correlated to a marked improvement in response and progression- free survival (57,59,63,67). It might be hypothesized that these immunomodulatory abilities have important implication for sustained control of the tumor and further deepening of response (63). As a

result of these pleiotropic immune modulation, CD38 antibodies also enhance anti-tumor activity of

others anti-cancer drugs with several studies highlighting that CD38-targeting antibodies have strong

synergistic activity, such as combination to lenalidomide as well as to PD1/PD-L1 inhibitors (56,68).

 Besides effect on immune cells, CD38 antibodies may also modulate immunometabolic pathway. Indeed, CD38-targeting agent's exposure could lead to lower adenosine level in tumoral

microenvironment, which is known as immunosuppressive metabolite (69,70). All these properties

enhancing the anti-tumoral response are of major interest in the field of oncology while it could be

problematic in immunosuppressive strategies such as autoimmune diseases treatment or

desensitization and SOT's context.

CD38 antibodies in solid organ transplantation

CD38 antibodies in non tumoral context

 In the last decade, several strategies to handle with autoimmune or alloimmune pathologic situations include CD38 antibodies (71–73). Indeed, long-lived plasma cells, which produce pathogenic antibodies, are unresponsive to standard immunosuppression. Besides PC depletion and immunomodulatory effect, CD38 expression on PCs from patients with autoimmune condition (74) and reduction of auto-antibodies in patients exposed to daratumumab (75) support the evaluation of daratumumab in patients with autoantibody-dependent disorders and, in extension, to alloimmune situation such as SOT. Available evidence about CD38 antibodies efficacy in these situations are mostly cases reports of daratumumab use against immune cytopenia. Daratumumab were used to treat warm autoimmune hemolytic anemia post-hematopoietic stem cell transplant (76), refractory 180 cold agglutinin disease (77), Evans syndrome (78) and pure red cell aplasia (79) with improvement in the majority of cases. Regarding other autoimmune disease, the administration of daratumumab in the majority of cases. Regarding other autoimmune disease, the administration of daratumumab in two patients with refractory lupus was recently described exhibiting clinical responses associated with significant depletion of long-lived plasma cells and modulation of effector T-cell responses (80). As regard as autoimmune encephalitis, targeting CD38 was achieved with daratumumab in one case of life-threatening anti-NMDA receptor encephalitis and in one case of refractory anti-CASPR2 encephalitis with improvements of neurological sequelae (81,82). In the last case, severe septicemia leading to patient death highlight an unmet need of rigorous clinical investigation to determine the efficacy and tolerance of CD38-targeting agent in autoimmune disease.

CD38 antibodies and ABMR treatment

 In antibody-mediated non-neoplastic diseases, alloimmune situation such as SOT represent a field where targeting CD38 is promising. As alloantibody-producing PC express CD38 at a higher 193 level than other CD38⁺ hematopoietic cells and CD38 antibodies induce a profound depletion of 194 CD38⁺ PC, CD38 appears as a rational target to handle with harmful alloantibodies such as DSA (83,84). Currently, only few studies have been published regarding the use of CD38 antibodies for desensitization in patients awaiting transplantation or for treatment of ABMR. Concerning treatment of ABMR, the first report was in a patient with refractory early active ABMR caused by anti-A isohemagglutinins after kidney transplantation from his ABO-incompatible sister (85) . Based on the efficacy of daratumumab in the treatment of pure red cell aplasia following ABO-incompatible hematopoietic stem cell (79) and non-response of several therapies; daratumumab were tested as a rescue solution leading to a significant decrease of the pathogenic isohemagglutinins and resolution of tissue damage in the kidney biopsy. Kwun and colleagues also published a case report of daratumumab as a therapeutic strategy for refractory heart and kidney rejection in a patient who received heart and kidney transplants due to systemic lupus (72). Both transplant biopsy showed T cell–mediated rejection, ABMR and diffuse PC infiltration associated to the presence of several DSA. To face refractory cardiogenic shock and acute kidney failure dependent to dialysis, a compassionate use of daratumumab lead to the resolution of both allograft function, improvement in acute kidney lesions with decreased PCs infiltrate and dramatic decline for the majority of DSA. A recurrent acute PC-rich rejection on kidney biopsy and significant ascension of DSA were successfully managed with daratumumab. Recently, two others cases were reported: one refractory ABMR after a heart transplant successfully treated with daratumumab and one chronic active ABMR in a kidney allograft recipient diagnosed with myeloma exposed to daratumumab (73,86). In the last one, the exhaustive immuno-monitoring showed that the main mode of action seems to be based on PC depletion, with profound PCs reduction in the bone marrow and peripheral blood and the abrogation of in vitro alloantibody production by PC enriched from bone marrow aspirates, leading to significant reduction in DSA levels (73). Another observation is that daratumumab led to depletion

 of NK cells infiltrating the allograft and circulating NK cells, which is major interest knowing the potential role of NK cells in microvasculature inflammation through engagement of their Fc gamma receptor IIIA with endothelium-bound DSA (87). Interestingly, while follow up biopsy showed resolution of humoral activity, it was observed tubulointerstitial inflammation which prompted steroid treatment. The author highlighted that the molecular signature of this infiltrate was not similar to signature of T-cell mediated rejection leading to question the trigger of this infiltrate not associated with graft dysfunction. Indeed, daratumumab may trigger T-cell alloresponse, even if circulating regulatory T cells were not reduced in the patient's blood which is not necessarily correlated to the modification of immune cell populations at a tissue level. Moreover, the authors recently reported long term data of this case without evidence of ABMR rebound after daratumumab discontinuation (88). Although it is difficult to decipher the role of a rescue with daratumumab added to a complex antirejection therapy, a drug that specifically deplete PC with a favorable safety profile could represent a step forward in the field.

CD38 antibodies and desensitization

 The ability of CD38 to desensitize has been evaluated in both preclinical and clinical contexts and published in the same study (72). The preclinical study was based on the use of daratumumab in a non-human primate model which has the most biological similarity to humans for solid organ transplant biology (41,89). The authors paired donors and recipients for maximal HLA mismatching and practiced, for allosensitization, two serial skin grafts before transplantation with a kidney from paired skin graft donor (72). Daratumumab and plerixafor (anti‐ CXCR4), known to induce mobilization of PC from bone marrow to peripheral blood, were given as desensitization therapy with an initiation 8-12 weeks after sensitization and 8 weeks before kidney transplantation. Animals received for induction anti-CD4 and anti-CD8 antibodies and for maintenance immunosuppression tacrolimus, mycophenolate mofetil and a methylprednisolone taper. This desensitization regimen reduced significantly preformed DSA, with more than 50% reduction compared with the pretreatment time point, and prolonged graft survival with a depletion of PC without altering the germinal center response since the Tfh population was not eliminated (72). However, desensitized monkeys showed delayed ABMR associated to DSA rebound and T cell–mediated rejection perhaps due to immune deviation. Indeed, the authors observed a reduction of regulatory B and T cells after desensitization with rapid emergence of activated T cells after kidney transplantation. This observation could be related to immunomodulatory effects of daratumumab but CXCR4 inhibition, due to plerixafor, is also known to limit regulatory compartment and to promote effector cells with a potential role in these cell‐ mediated rejection (90). Thus, in transplant recipients following desensitization with 251 daratumumab, it would be interesting to elaborate new strategies than current immunosuppressive
252 regimens in order to manage these DSA rebounds and the risk of T cell–mediated rejection. regimens in order to manage these DSA rebounds and the risk of T cell–mediated rejection. Concerning the clinical setting, the authors used daratumumab in a heart transplant candidate remaining highly sensitized after multiple courses of plasmapheresis, high-dose IVIG, and rituximab. It was observed a significant and persistent decrease of allosensitization allowing a heart transplantation six months after daratumumab infusion (72). Currently, based on these promising results, daratumumab are under investigation for desensitization in patients awaiting solid-organ transplantation in two clinical trial, one ruled by the nephrology department of Henri Mondor Hospital (Créteil, France) and another one directed by Stanford University (ClinicalTrials.gov, NCT04204980 and NCT04088903 (91,92)). Regarding the trial in kidney transplantation, sensitized patients with calculated panel reactive antibodies (cPRA) > 95% awaiting on the French National kidney allograft waiting-list for at least three years are eligible for the study and are randomly assigned to one of the two steps : (step 1) dose-escalation with 4 mg/kg of daratumumab weekly for four weeks, then with 8 mg/kg weekly for four weeks and then 16 mg/kg weekly for four weeks; (step 2) expansion cohort with eight weekly doses of 16 mg/kg. The primary outcomes are defined

- 266 as: adverse events, intra-patient variation of cPRA and anti-HLA levels. Several other outcomes are also of interest such as percentage of patients engrafted, and intra-patient variation of ABO antibody titers (91).
- also of interest such as percentage of patients engrafted, and intra-patient variation of ABO antibody
- 268 titers (91)**.**

Conclusion

 Therapeutic improvement is required for both prevention and treatment of humoral alloresponse in solid organ transplantation. CD38 antibodies are a promising solution to profoundly 273 deplete high affinity anti-HLA producing plasma cells. Preclinical and clinical experimental results
274 suggests that daratumumab is a potentially therapeutic strategy to reduce DSA production and suggests that daratumumab is a potentially therapeutic strategy to reduce DSA production and prevent and/or treat antibody-mediated rejection. However, CD38-targeting agent induce immune 276 deviation which could be deleterious for solid organ transplants enhancing cellular-mediated
277 rejection. Clinical studies are now needed to clarify the indications and efficacy of these promising rejection. Clinical studies are now needed to clarify the indications and efficacy of these promising

- therapeutic strategies.
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References

- 1. OPTN: Organ Procurement and Transplantation Network OPTN [Internet]. [cited 2021 Mar 14]. Available from: https://optn.transplant.hrsa.gov/
- 2. Agence de la biomédecine [Internet]. [cited 2021 Mar 14]. Available from:
- https://www.agence-biomedecine.fr/
- 3. Loupy A, Lefaucheur C. Antibody-Mediated Rejection of Solid-Organ Allografts. N Engl J
- Med [Internet]. 2018 Sep 19 [cited 2021 Mar 14]; Available from:
- https://www.nejm.org/doi/10.1056/NEJMra1802677
- 4. Valenzuela NM, Reed EF. Antibody-mediated rejection across solid organ transplants:
- manifestations, mechanisms, and therapies. J Clin Invest. 2017 Jun 30;127(7):2492–504.
- 5. The XIIIth Banff Conference on Allograft Pathology: The Banff 2015 Heart Meeting Report:
- Improving Antibody-Mediated Rejection Diagnostics: Strengths, Unmet Needs, and Future
- 292 Directions PubMed [Internet]. [cited 2021 Mar 14]. Available from:
293 https://pubmed.ncbi.nlm.nih.gov/27862968/
- https://pubmed.ncbi.nlm.nih.gov/27862968/
- 6. Schinstock CA, Askar M, Bagnasco SM, Batal I, Bow L, Budde K, et al. A 2020 Banff
- Antibody-mediatedInjury Working Group examination of international practices for diagnosing
- antibody-mediated rejection in kidney transplantation a cohort study. Transpl Int Off J Eur Soc Organ Transplant. 2021 Mar;34(3):488–98.
- 7. Roux A, Levine DJ, Zeevi A, Hachem R, Halloran K, Halloran PF, et al. Banff Lung Report: Current knowledge and future research perspectives for diagnosis and treatment of pulmonary
- antibody-mediated rejection (AMR). Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. 2019 Jan;19(1):21–31.
- 8. Drachenberg CB, Torrealba JR, Nankivell BJ, Rangel EB, Bajema IM, Kim DU, et al. Guidelines for the diagnosis of antibody-mediated rejection in pancreas allografts-updated Banff grading schema. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. 2011
- Sep;11(9):1792–802.
- 9. Demetris AJ, Bellamy C, Hübscher SG, O'Leary J, Randhawa PS, Feng S, et al. 2016 Comprehensive Update of the Banff Working Group on Liver Allograft Pathology: Introduction of Antibody-Mediated Rejection. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. 2016 Oct;16(10):2816–35.
- 10. Loupy A, Haas M, Roufosse C, Naesens M, Adam B, Afrouzian M, et al. The Banff 2019 Kidney Meeting Report (I): Updates on and clarification of criteria for T cell- and antibody-mediated
- rejection. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. 2020 Sep;20(9):2318–31.
- 11. Zhang R. Donor-Specific Antibodies in Kidney Transplant Recipients. Clin J Am Soc
- Nephrol CJASN. 2018 Jan 6;13(1):182–92.
- 12. Butler CL, Valenzuela NM, Thomas KA, Reed EF. Not All Antibodies Are Created Equal: Factors That Influence Antibody Mediated Rejection. J Immunol Res. 2017;2017:7903471.
- 13. Lefaucheur C, Viglietti D, Mangiola M, Loupy A, Zeevi A. From Humoral Theory to
- Performant Risk Stratification in Kidney Transplantation. J Immunol Res. 2017;2017:5201098.
- 14. Lefaucheur C, Viglietti D, Bentlejewski C, Duong van Huyen J-P, Vernerey D, Aubert O, et
- al. IgG Donor-Specific Anti-Human HLA Antibody Subclasses and Kidney Allograft Antibody-
- Mediated Injury. J Am Soc Nephrol JASN. 2016 Jan;27(1):293–304.
- 15. Loupy A, Lefaucheur C, Vernerey D, Prugger C, Duong van Huyen J-P, Mooney N, et al. Complement-binding anti-HLA antibodies and kidney-allograft survival. N Engl J Med. 2013 Sep 26;369(13):1215–26.
- 16. Montgomery RA, Tatapudi VS, Leffell MS, Zachary AA. HLA in transplantation. Nat Rev Nephrol. 2018 Sep;14(9):558–70.
- 17. Tinckam KJ, Keshavjee S, Chaparro C, Barth D, Azad S, Binnie M, et al. Survival in
- sensitized lung transplant recipients with perioperative desensitization. Am J Transplant Off J Am
- Soc Transplant Am Soc Transpl Surg. 2015 Feb;15(2):417–26.
- 18. Plazak ME, Gale SE, Reed BN, Hammad S, Ton V-K, Kaczorowski DJ, et al. Clinical
- Outcomes of Perioperative Desensitization in Heart Transplant Recipients. Transplant Direct. 2021 Feb;7(2):e658.
- 19. Bourassa-Blanchette S, Patel V, Knoll GA, Hutton B, Fergusson N, Bennett A, et al. Clinical outcomes of polyvalent immunoglobulin use in solid organ transplant recipients: A systematic review
- and meta-analysis Part II: Non-kidney transplant. Clin Transplant. 2019 Jul;33(7):e13625.
- 20. Shah KS, Patel J. Desensitization in heart transplant recipients: Who, when, and how. Clin Transplant. 2019 Aug;33(8):e13639.
- 21. Matsumoto CS, Rosen-Bronson S. Donor-specific antibody and sensitized patients in intestinal transplantation. Curr Opin Organ Transplant. 2021 Apr 1;26(2):245–9.
- 22. Schinstock CA, Smith BH, Montgomery RA, Jordan SC, Bentall AJ, Mai M, et al. Managing highly sensitized renal transplant candidates in the era of kidney paired donation and the new kidney allocation system: Is there still a role for desensitization? Clin Transplant. 2019 Dec;33(12):e13751.
- 23. Vo AA, Choi J, Cisneros K, Reinsmoen N, Haas M, Ge S, et al. Benefits of rituximab
- combined with intravenous immunoglobulin for desensitization in kidney transplant recipients.
- Transplantation. 2014 Aug 15;98(3):312–9.
- 24. Vo AA, Peng A, Toyoda M, Kahwaji J, Cao K, Lai C-H, et al. Use of intravenous immune globulin and rituximab for desensitization of highly HLA-sensitized patients awaiting kidney transplantation. Transplantation. 2010 May 15;89(9):1095–102.
- 25. Vo AA, Lukovsky M, Toyoda M, Wang J, Reinsmoen NL, Lai C-H, et al. Rituximab and intravenous immune globulin for desensitization during renal transplantation. N Engl J Med. 2008 Jul 17;359(3):242–51.
- 26. Loupy A, Suberbielle-Boissel C, Zuber J, Anglicheau D, Timsit M-O, Martinez F, et al.
- Combined posttransplant prophylactic IVIg/anti-CD 20/plasmapheresis in kidney recipients with
- preformed donor-specific antibodies: a pilot study. Transplantation. 2010 Jun 15;89(11):1403–10.
- 27. Jordan SC, Toyoda M, Kahwaji J, Vo AA. Clinical Aspects of Intravenous Immunoglobulin Use in Solid Organ Transplant Recipients. Am J Transplant. 2011;11(2):196–202.
- 28. Amrouche L, Aubert O, Suberbielle C, Rabant M, Van Huyen J-PD, Martinez F, et al. Long-term Outcomes of Kidney Transplantation in Patients With High Levels of Preformed DSA: The
- Necker High-Risk Transplant Program. Transplantation. 2017 Oct;101(10):2440–8.
- 29. Orandi BJ, Luo X, King EA, Garonzik-Wang JM, Bae S, Montgomery RA, et al. Hospital
- readmissions following HLA-incompatible live donor kidney transplantation: A multi-center study. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. 2018 Mar;18(3):650–8.
- 30. Motter JD, Jackson KR, Long JJ, Waldram MM, Orandi BJ, Montgomery RA, et al. Delayed
- graft function and acute rejection following HLA-incompatible living donor kidney transplantation. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. 2020 Dec 28;
- 31. Kahwaji J, Jordan SC, Najjar R, Wongsaroj P, Choi J, Peng A, et al. Six-year outcomes in
- broadly HLA-sensitized living donor transplant recipients desensitized with intravenous
- immunoglobulin and rituximab. Transpl Int Off J Eur Soc Organ Transplant. 2016 Dec;29(12):1276– 85.
- 32. Süsal C, Opelz G. Transplantation: Desensitization and survival in kidney transplant recipients. Nat Rev Nephrol. 2017 Apr;13(4):196–8.
- 33. Heidt S, Claas FHJ. Transplantation in highly sensitized patients: challenges and
- recommendations. Expert Rev Clin Immunol. 2018 Aug 3;14(8):673–9.
- 34. Montgomery RA, Lonze BE, King KE, Kraus ES, Kucirka LM, Locke JE, et al.
- Desensitization in HLA-incompatible kidney recipients and survival. N Engl J Med. 2011 Jul 28;365(4):318–26.
- 35. Orandi BJ, Luo X, Massie AB, Garonzik-Wang JM, Lonze BE, Ahmed R, et al. Survival
- Benefit with Kidney Transplants from HLA-Incompatible Live Donors. N Engl J Med. 2016 Mar 10;374(10):940–50.
- 36. Manook M, Koeser L, Ahmed Z, Robb M, Johnson R, Shaw O, et al. Post-listing survival for highly sensitised patients on the UK kidney transplant waiting list: a matched cohort analysis. Lancet Lond Engl. 2017 18;389(10070):727–34.
- 37. Jordan SC, Ammerman N, Choi J, Huang E, Peng A, Sethi S, et al. The role of novel
- therapeutic approaches for prevention of allosensitization and antibody-mediated rejection. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. 2020 Jun;20 Suppl 4:42–56.
- 38. Leibler C, Thiolat A, Elsner RA, El Karoui K, Samson C, Grimbert P. Costimulatory
- blockade molecules and B-cell-mediated immune response: current knowledge and perspectives. Kidney Int. 2019 Apr;95(4):774–86.
- 39. Tangye SG, Ma CS, Brink R, Deenick EK. The good, the bad and the ugly TFH cells in 390 human health and disease. Nat Rev Immunol. 2013 Jun;13(6):412–26.
391 40. Nutt SL, Hodgkin PD, Tarlinton DM, Corcoran LM. The gener
- 40. Nutt SL, Hodgkin PD, Tarlinton DM, Corcoran LM. The generation of antibody-secreting plasma cells. Nat Rev Immunol. 2015 Mar;15(3):160–71.
- 41. Kwun J, Knechtle S. Experimental modeling of desensitization: What have we learned about
- preventing AMR? Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. 2020 Jun;20 Suppl 4:2–11.
- 42. Brynjolfsson SF, Persson Berg L, Olsen Ekerhult T, Rimkute I, Wick M-J, Mårtensson I-L, et al. Long-Lived Plasma Cells in Mice and Men. Front Immunol. 2018;9:2673.
- 43. Jordan SC, Ammerman N, Choi J, Kumar S, Huang E, Toyoda M, et al. Interleukin-6: An Important Mediator of Allograft Injury. Transplantation. 2020 Dec;104(12):2497–506.
- 44. Choi J, Aubert O, Vo A, Loupy A, Haas M, Puliyanda D, et al. Assessment of Tocilizumab
- (Anti-Interleukin-6 Receptor Monoclonal) as a Potential Treatment for Chronic Antibody-Mediated
- Rejection and Transplant Glomerulopathy in HLA-Sensitized Renal Allograft Recipients. Am J
- Transplant Off J Am Soc Transplant Am Soc Transpl Surg. 2017 Sep;17(9):2381–9.
- 45. Clazakizumab as an Agent to Reduce Donor Specific HLA Antibodies and Improve
- Outcomes in Patients with Chronic & Active Antibody-Mediated Rejection Post-Kidney
- Transplantation [Internet]. ATC Abstracts. [cited 2021 Mar 21]. Available from:
- https://atcmeetingabstracts.com/abstract/clazakizumab-as-an-agent-to-reduce-donor-specific-hla-
- antibodies-and-improve-outcomes-in-patients-with-chronic-active-antibody-mediated-rejection-post-kidney-transplantation/
- 46. MD SJ. A Phase I/II Trial to Evaluate the Safety and Tolerability of Clazakizumab (Anti-IL-6
- Monoclonal) to Eliminate Donor Specific HLA Antibodies (DSAs) and Improve Transplant Rates in
- Highly-HLA Sensitized Patients Awaiting Renal Transplant [Internet]. clinicaltrials.gov; 2020 Oct
- [cited 2021 Mar 18]. Report No.: NCT03380962. Available from:
- https://clinicaltrials.gov/ct2/show/NCT03380962
- 47. Woodle ES, Shields AR, Ejaz NS, Sadaka B, Girnita A, Walsh RC, et al. Prospective Iterative
- Trial of Proteasome Inhibitor-Based Desensitization. Am J Transplant. 2015;15(1):101–18.
- 48. Tremblay S, Driscoll JJ, Rike-Shields A, Hildeman DA, Alloway RR, Girnita AL, et al. A
- prospective, iterative, adaptive trial of carfilzomib-based desensitization. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. 2020 Feb;20(2):411–21.
- 49. Kwun J, Burghuber C, Manook M, Iwakoshi N, Gibby A, Hong JJ, et al. Humoral
- Compensation after Bortezomib Treatment of Allosensitized Recipients. J Am Soc Nephrol JASN. 2017 Jul;28(7):1991–6.
- 50. Ezekian B, Schroder PM, Mulvihill MS, Barbas A, Collins B, Freischlag K, et al.
- Pretransplant Desensitization with Costimulation Blockade and Proteasome Inhibitor Reduces DSA
- and Delays Antibody-Mediated Rejection in Highly Sensitized Nonhuman Primate Kidney
- Transplant Recipients. J Am Soc Nephrol JASN. 2019 Dec;30(12):2399–411.
- 51. Schroder PM, Schmitz R, Fitch ZW, Ezekian B, Yoon J, Choi AY, et al. Preoperative
- carfilzomib and lulizumab based desensitization prolongs graft survival in a sensitized non-human primate model. Kidney Int. 2021 Jan;99(1):161–72.
- 52. Quarona V, Zaccarello G, Chillemi A, Brunetti E, Singh VK, Ferrero E, et al. CD38 and
- CD157: A long journey from activation markers to multifunctional molecules. Cytometry B Clin Cytom. 2013;84B(4):207–17.
- 53. Dianzani U, Malavasi F. Lymphocyte adhesion to endothelium. Crit Rev Immunol. 1995;15(2):167–200.
- 54. van de Donk NWCJ, Usmani SZ. CD38 Antibodies in Multiple Myeloma: Mechanisms of Action and Modes of Resistance. Front Immunol. 2018;9:2134.
- 55. van de Donk NWCJ, Richardson PG, Malavasi F. CD38 antibodies in multiple myeloma: back to the future. Blood. 2018 Jan 4;131(1):13–29.
- 56. van de Donk NWCJ, Pawlyn C, Yong KL. Multiple myeloma. Lancet Lond Engl. 2021 Jan 30;397(10272):410–27.
- 57. Lokhorst HM, Plesner T, Laubach JP, Nahi H, Gimsing P, Hansson M, et al. Targeting CD38 with Daratumumab Monotherapy in Multiple Myeloma. N Engl J Med. 2015 Sep 24;373(13):1207– 19.
- 58. Casneuf T, Xu XS, Adams HC, Axel AE, Chiu C, Khan I, et al. Effects of daratumumab on
- natural killer cells and impact on clinical outcomes in relapsed or refractory multiple myeloma. Blood Adv. 2017 Oct 24;1(23):2105–14.
- 59. Dimopoulos MA, Oriol A, Nahi H, San-Miguel J, Bahlis NJ, Usmani SZ, et al.
- Daratumumab, Lenalidomide, and Dexamethasone for Multiple Myeloma. N Engl J Med. 2016 Oct 6;375(14):1319–31.
- 60. Touzeau C, Moreau P. Daratumumab for the treatment of multiple myeloma. Expert Opin Biol Ther. 2017;17(7):887–93.
- 61. Lammerts van Bueren J, Jakobs D, Kaldenhoven N, Roza M, Hiddingh S, Meesters J, et al.
- Direct in Vitro Comparison of Daratumumab with Surrogate Analogs of CD38 Antibodies
- MOR03087, SAR650984 and Ab79. Blood. 2014 Dec 6;124(21):3474–3474.
- 62. Radocha J, van de Donk NWCJ, Weisel K. Monoclonal Antibodies and Antibody Drug Conjugates in Multiple Myeloma. Cancers. 2021 Mar 29;13(7).
- 63. van de Donk NWCJ. Immunomodulatory effects of CD38-targeting antibodies. Immunol Lett. 2018 Jul;199:16–22.
- 64. Krejcik J, Casneuf T, Nijhof IS, Verbist B, Bald J, Plesner T, et al. Daratumumab depletes
- CD38+ immune regulatory cells, promotes T-cell expansion, and skews T-cell repertoire in multiple myeloma. Blood. 2016 Jul 21;128(3):384–94.
- 65. Zhang L, Tai Y-T, Ho M, Xing L, Chauhan D, Gang A, et al. Regulatory B cell-myeloma cell
- interaction confers immunosuppression and promotes their survival in the bone marrow milieu.
- Blood Cancer J. 2017 Mar 24;7(3):e547.
- 66. Dwivedi S, Rendón-Huerta EP, Ortiz-Navarrete V, Montaño LF. CD38 and Regulation of the Immune Response Cells in Cancer. J Oncol. 2021;2021:6630295.
- 67. Adams HC, Stevenaert F, Krejcik J, Van der Borght K, Smets T, Bald J, et al. High-
- Parameter Mass Cytometry Evaluation of Relapsed/Refractory Multiple Myeloma Patients Treated
- with Daratumumab Demonstrates Immune Modulation as a Novel Mechanism of Action. Cytom Part
- J Int Soc Anal Cytol. 2019 Mar;95(3):279–89.
- 68. Chen L, Diao L, Yang Y, Yi X, Rodriguez BL, Li Y, et al. CD38-Mediated
- Immunosuppression as a Mechanism of Tumor Cell Escape from PD-1/PD-L1 Blockade. Cancer Discov. 2018 Sep;8(9):1156–75.
- 69. Vijayan D, Young A, Teng MWL, Smyth MJ. Targeting immunosuppressive adenosine in cancer. Nat Rev Cancer. 2017 Dec;17(12):709–24.

 70. Chatterjee S, Daenthanasanmak A, Chakraborty P, Wyatt MW, Dhar P, Selvam SP, et al. CD38-NAD+Axis Regulates Immunotherapeutic Anti-Tumor T Cell Response. Cell Metab. 2018 Jan 9;27(1):85-100.e8. 71. Zaninoni A, Giannotta JA, Gallì A, Artuso R, Bianchi P, Malcovati L, et al. The Immunomodulatory Effect and Clinical Efficacy of Daratumumab in a Patient With Cold Agglutinin Disease. Front Immunol. 2021;12:649441. 72. Kwun J, Matignon M, Manook M, Guendouz S, Audard V, Kheav D, et al. Daratumumab in Sensitized Kidney Transplantation: Potentials and Limitations of Experimental and Clinical Use. J Am Soc Nephrol JASN. 2019 Jul;30(7):1206–19. 73. Doberer K, Kläger J, Gualdoni GA, Mayer KA, Eskandary F, Farkash EA, et al. CD38 Antibody Daratumumab for the Treatment of Chronic Active Antibody-mediated Kidney Allograft Rejection. Transplantation. 2021 Feb 1;105(2):451–7. 488 74. Cole S, Walsh A, Yin X, Wechalekar MD, Smith MD, Proudman SM, et al. Integrative
489 analysis reveals CD38 as a therapeutic target for plasma cell-rich pre-disease and established analysis reveals CD38 as a therapeutic target for plasma cell-rich pre-disease and established rheumatoid arthritis and systemic lupus erythematosus. Arthritis Res Ther. 2018 May 2;20(1):85. 75. Frerichs KA, Verkleij CPM, Bosman PWC, Zweegman S, Otten H, van de Donk NWCJ. CD38-targeted therapy with daratumumab reduces autoantibody levels in multiple myeloma patients. J Transl Autoimmun. 2019 Dec;2:100022. 76. Tolbert VP, Goldsby R, Huang J, Shimano K, Melton A, Willert J, et al. Daratumumab Is Effective in the Treatment of Refractory Post-Transplant Autoimmune Hemolytic Anemia: A Pediatric Case Report. Blood. 2016 Dec 2;128(22):4819–4819. 77. Tomkins O, Berentsen S, Arulogun S, Sekhar M, D'Sa S. Daratumumab for disabling cold agglutinin disease refractory to B-cell directed therapy. Am J Hematol. 2020 Jul 11; 78. Blennerhassett R, Sudini L, Gottlieb D, Bhattacharyya A. Post-allogeneic transplant Evans syndrome successfully treated with daratumumab. Br J Haematol. 2019 Oct;187(2):e48–51. 79. Chapuy CI, Kaufman RM, Alyea EP, Connors JM. Daratumumab for Delayed Red-Cell Engraftment after Allogeneic Transplantation. N Engl J Med. 2018 08;379(19):1846–50. 80. Ostendorf L, Burns M, Durek P, Heinz GA, Heinrich F, Garantziotis P, et al. Targeting CD38 with Daratumumab in Refractory Systemic Lupus Erythematosus. N Engl J Med. 2020 Sep 17;383(12):1149–55. 81. Ratuszny D, Skripuletz T, Wegner F, Groß M, Falk C, Jacobs R, et al. Case Report: Daratumumab in a Patient With Severe Refractory Anti-NMDA Receptor Encephalitis. Front Neurol. 2020;11:602102. 82. Scheibe F, Ostendorf L, Reincke SM, Prüss H, von Brünneck A-C, Köhnlein M, et al. Daratumumab treatment for therapy-refractory anti-CASPR2 encephalitis. J Neurol. 2020 Feb;267(2):317–23. 83. Martin TG, Corzo K, Chiron M, Velde H van de, Abbadessa G, Campana F, et al. Therapeutic Opportunities with Pharmacological Inhibition of CD38 with Isatuximab. Cells. 2019 Nov 26;8(12). 84. Flores-Montero J, de Tute R, Paiva B, Perez JJ, Böttcher S, Wind H, et al. Immunophenotype of normal vs. myeloma plasma cells: Toward antibody panel specifications for MRD detection in multiple myeloma. Cytometry B Clin Cytom. 2016 Jan;90(1):61–72. 85. Spica D, Junker T, Dickenmann M, Schaub S, Steiger J, Rüfli T, et al. Daratumumab for Treatment of Antibody-Mediated Rejection after ABO-Incompatible Kidney Transplantation. Case Rep Nephrol Dial. 2019 Dec;9(3):149–57. 86. Aguilera Agudo C, Gómez Bueno M, Krsnik Castello I. Daratumumab for Antibody- mediated Rejection in Heart Transplant-A Novel Therapy: Successful Treatment of Antibody- mediated Rejection. Transplantation. 2021 Mar 1;105(3):e30–1. 87. Parkes MD, Halloran PF, Hidalgo LG. Evidence for CD16a-Mediated NK Cell Stimulation in

- Antibody-Mediated Kidney Transplant Rejection. Transplantation. 2017 Apr;101(4):e102–11.
- 88. Mayer KA, Doberer K, Eskandary F, Halloran PF, Böhmig GA. New concepts in chronic
- antibody-mediated kidney allograft rejection: prevention and treatment. Curr Opin Organ Transplant. 2021 Feb 1;26(1):97–105.
-
- 89. Kirk AD. Crossing the bridge: large animal models in translational transplantation research.
- Immunol Rev. 2003 Dec;196:176–96.
- 90. Mollica Poeta V, Massara M, Capucetti A, Bonecchi R. Chemokines and Chemokine
- Receptors: New Targets for Cancer Immunotherapy. Front Immunol. 2019;10:379.
- 91. Assistance Publique Hôpitaux de Paris. Desensitization in Kidney Allograft Recipients
- Before Transplantation Using Daratumumab [Internet]. clinicaltrials.gov; 2020 May [cited 2021 Mar
- 23]. Report No.: NCT04204980. Available from: https://clinicaltrials.gov/ct2/show/NCT04204980
- 92. Witteles R. A Phase 1 Study of Daratumumab for Reduction of Circulating Antibodies in
- Patients With High Allosensitization Awaiting Heart Transplantation [Internet]. clinicaltrials.gov;
- 2020 Oct [cited 2021 Mar 23]. Report No.: NCT04088903. Available from:
- https://clinicaltrials.gov/ct2/show/NCT04088903

541 **Tables**

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543 **Table 1. Selection of therapeutical regimens targeting CD38**

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547 **Table 2. CD38 antibody use in solid organ transplantation**

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549 *ABMR: antibody mediated rejection, ATG: anti-human thymocytes globulins, DSA: donor specific*

550 *antibodies, IVIG: intravenous immunoglobulins, MMF: mycophenolate mofetil, NHP: non-human* 551 *primate, PC: plasma cells, Ref. : reference, TCMR: T cell mediated rejection, Tx: transplantation*

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- **Figures**
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- **Figure 1. Immune effects of anti-CD38 antibody in the context of solid organ transplantation.**
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- *ABMR: antibody mediated rejection, Breg: regulatory B cell, DSA: donor specific antibodies, PC:*
- *plasma cell, TCMR: T cell mediated rejection, Treg: regulatory T cell.*

1 Conflict of Interest

563 The authors declare that the review was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest. relationships that could be construed as a potential conflict of interest.

2 Author Contributions

566 NJ, MM and PG designed the review, collected and interpreted data from literature, and wrote the manuscript. manuscript.

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